

Boc-tyrosyl-D-alanyl-glycyl-N-methyl-phenylalanyl-O-methyl-methionine hydrate: a protected analog of metkephamid

Jeffrey R. Deschamps,^{a*} Judith L. Flippen-Anderson,^a George A. Brine,^b James P. Hayes^b and Clifford George^a

^aLaboratory for the Structure of Matter, Naval Research Laboratory, Washington, DC 20375, USA, and ^bChemistry and Life Sciences, Research Triangle Institute, Research Triangle Park, NC 27709, USA

Correspondence e-mail: deschamps@nrl.navy.mil

Key indicators

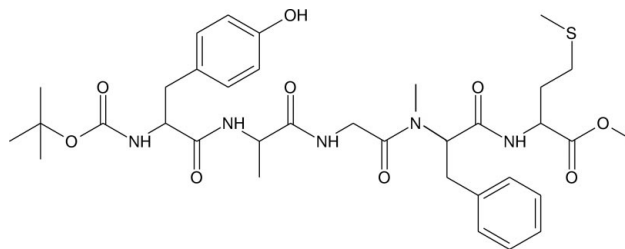
Single-crystal X-ray study
 T = 153 K
 Mean $\sigma(\text{C}-\text{C}) = 0.005 \text{ \AA}$
 Disorder in solvent or counterion
 R factor = 0.043
 wR factor = 0.112
 Data-to-parameter ratio = 10.9

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

In this study, we report on the X-ray diffraction analysis of the pentapeptide, Boc-Tyr-D-Ala-Gly-(NMe-Phe)-(Met-O-Me), $\text{C}_{35}\text{H}_{49}\text{N}_5\text{O}_9\text{S} \cdot 1.125\text{H}_2\text{O}$. This is a protected intermediate in the solution phase synthesis of metkephamid (LY127,623), Tyr-D-Ala-Gly-NMe-Phe-Met-NH₂. The peptide crystallizes in the monoclinic space group *C*2 with two peptide molecules and three solvent (water) molecules in the asymmetric unit. The peptide has an extended conformation similar to other *Met*-enkephalins.

Comment

Theoretically, linear peptides can have many different backbone conformations. Early X-ray studies (1983–1987) on enkephalin and its analogs showed only two different backbone conformations: extended and single β -bend. No other conformations were observed until a double β -bend was reported in 1989 (for a review, see Deschamps *et al.*, 1996). Despite the small number of conformations observed in the solid state, most enkephalin analogs can adopt many conformations in solution. To constrain the peptide conformation a number of modifications have been investigated, including the substitution, deletion or addition of natural or artificial amino acids and the formation of highly constrained peptide ring systems. In this study, we report on the X-ray diffraction analysis of the pentapeptide, Boc-Tyr-D-Ala-Gly-(NMe-Phe)-(Met-O-Me), a protected intermediate in the solution phase synthesis of metkephamid (LY127,623). Metkephamide (Tyr-D-Ala-Gly-NMe-Phe-Met-NH₂) is an analog of methionine-enkephalin that retains high affinity for the delta receptor, and has systemic analgesic activity (Frederickson *et al.*, 1981).



(I)

The title compound, (I), crystallizes with two peptide molecules and three water molecules in the asymmetric unit (Fig. 1). Additional disordered water molecules are probably present in the crystal, as evidenced by the diffuse residual electron density, and voids large enough to accommodate a water molecule were detected by *PLATON* (Spek, 2001). The two independent peptides molecules have an extended

Received 20 November 2001

Accepted 29 November 2001

Online 8 December 2001

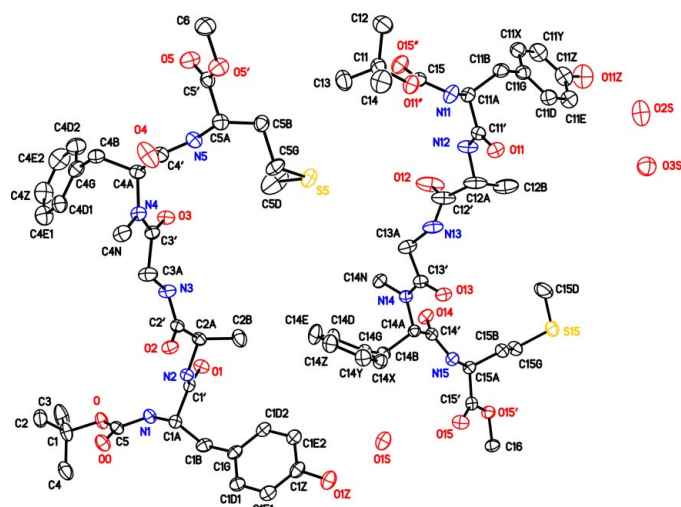


Figure 1
The structure of Boc-Tyr-D-Ala-Gly-NMe-Phe-Met-OMe showing the relationship of the two independent peptide molecules and the associated water molecules. Displacement ellipsoids are shown at the 30% probability level.

conformation, although even the main-chain atoms cannot be fully superimposed (Fig. 2). The extended conformation observed in this study is similar to that of other *Met*-enkephalins (Doi *et al.*, 1984; Griffin *et al.*, 1986; Doi *et al.*, 1987).

The peptide molecules form an extensive hydrogen-bonded network (Table 1). Pairs of adjacent peptide molecules are linked across the chains by six hydrogen bonds (Fig. 3). These pairs are then linked to adjacent pairs by two hydrogen bonds forming extended ribbons. This pattern of cross-chain hydrogen bonds is typical of enkephalins with an extended conformation (Karle *et al.*, 1983; Griffin *et al.*, 1986; Doi *et al.*, 1987). Two of the solvent (*i.e.* water) molecules form hydrogen

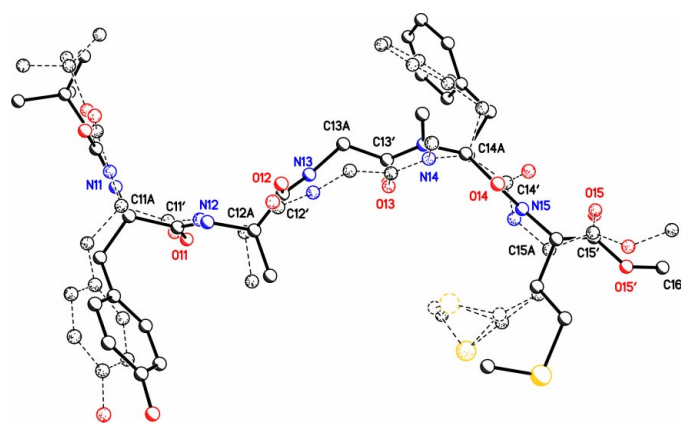


Figure 2
The conformation of Boc-Tyr-D-Ala-Gly-NMe-Phe-Met-OMe. The two independent peptide molecules are superimposed (alignment was based on the positions of the main-chain N and O atoms). Although the two independent molecules have similar conformations, even the main-chain atoms cannot be superimposed. Additional differences are present in the orientations of the side chains.

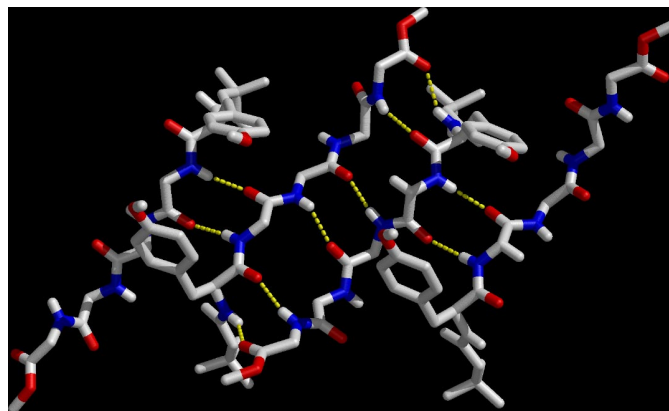


Figure 3
Inter-chain hydrogen bonding in Boc-Tyr-D-Ala-Gly-NMe-Phe-Met-OMe. The six hydrogen bonds between the pair of peptide molecules in the asymmetric unit and the two hydrogen bonds connecting adjacent pairs are shown as dashed lines. Solvent molecules and side chains not involved in hydrogen bonding have been omitted for clarity.

bonds between adjacent ribbons; the third water molecule (O3S) interacts only with other solvent molecules.

In this study, the X-ray diffraction data were collected using a Bruker SMART 6000 detector and a copper rotating anode. Prior to this, we collected X-ray diffraction data at the Cornell High Energy Synchrotron Source (CHESS). The structure determined from the synchrotron data is identical to that reported here. However, the synchrotron determined structure suffered from an incomplete dataset (due to the experimental set up) and unusually large sigmas for the measured intensities. Because of these problems the structure-solution for the synchrotron data was more difficult and the structure did not refine as well.

Experimental

The peptide, Boc-Tyr-D-Ala-Gly-NMe-Phe-Met-OMe, was supplied by Research Triangle Institute and was prepared using standard methods of peptide synthesis. Crystals were grown by vapor diffusion of petroleum ether into acetone.

Crystal data

$C_{35}H_{49}N_5O_9S \cdot 1.125H_2O$
 $M_r = 735.87$
 Monoclinic, $C2$
 $a = 38.354 (1) \text{ \AA}$
 $b = 11.000 (1) \text{ \AA}$
 $c = 22.689 (1) \text{ \AA}$
 $\beta = 123.15 (1)^\circ$
 $V = 8014.4 (8) \text{ \AA}^3$
 $Z = 8$

$D_x = 1.220 \text{ Mg m}^{-3}$
 Cu $K\alpha$ radiation
 Cell parameters from 928 reflections
 $\theta = 5.5\text{--}124.6^\circ$
 $\mu = 1.21 \text{ mm}^{-1}$
 $T = 153 (2) \text{ K}$
 Prism, colorless
 $0.52 \times 0.50 \times 0.46 \text{ mm}$

Data collection

Bruker SMART 6000 CCD diffractometer
 ω scans
 Absorption correction: multi-scan (SADABS; Bruker, 2000)
 $T_{\min} = 0.559$, $T_{\max} = 0.573$
 17149 measured reflections

10567 independent reflections
 8802 reflections with $I > 2\sigma(I)$
 $R_{\text{int}} = 0.033$
 $\theta_{\text{max}} = 65.1^\circ$
 $h = -42 \rightarrow 44$
 $k = -12 \rightarrow 12$
 $l = -26 \rightarrow 25$

Refinement

Refinement on F^2
 $R[F^2 > 2\sigma(F^2)] = 0.043$
 $wR(F^2) = 0.112$
 $S = 1.01$
 10567 reflections
 966 parameters
 H atoms treated by a mixture of
 independent and constrained
 refinement

$w = 1/[\sigma^2(F_o^2) + (0.0576P)^2]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} = 0.056$
 $\Delta\rho_{\max} = 0.22 \text{ e } \text{\AA}^{-3}$
 $\Delta\rho_{\min} = -0.23 \text{ e } \text{\AA}^{-3}$
 Extinction correction: *SHELXL97*
 Extinction coefficient: 0.000290 (16)
 Absolute structure: Flack (1983)
 Flack parameter = 0.00 (2)

Table 1

Hydrogen-bonding geometry (\AA , $^\circ$).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
$N1-H1A\cdots O15$	0.86	2.29	3.036 (3)	145
$O1Z-H1ZA\cdots O1S$	0.82	1.89	2.708 (3)	179
$N2-H2D\cdots O2^i$	0.86	2.11	2.960 (3)	169
$N3-H3D\cdots O13$	0.86	2.26	3.029 (3)	149
$N5-H5A\cdots O11$	0.86	2.07	2.865 (3)	153
$N11-H11A\cdots O5$	0.86	2.26	3.017 (3)	146
$O11Z-H11G\cdots O2S^{ii}$	0.82	1.93	2.745 (5)	175
$N12-H12D\cdots O12^{iii}$	0.86	2.10	2.956 (4)	171
$N13-H13D\cdots O3$	0.86	2.05	2.902 (3)	169
$N15-H15A\cdots O1$	0.86	2.00	2.828 (3)	161
$O1S-H1SA\cdots O2^{iv}$	0.85 (4)	2.02 (4)	2.842 (3)	164 (3)
$O1S-H1SB\cdots O0^{ii}$	0.86 (3)	2.00 (3)	2.827 (3)	160 (4)
$O2S-H2SA\cdots O15^v$	0.89 (2)	1.93 (2)	2.821 (3)	175 (4)
$O2S-H2SB\cdots O3S^{iii}$	0.87 (2)	1.76 (3)	2.521 (14)	144 (4)
$O2S-H2SB\cdots O3S$	0.87 (2)	2.01 (3)	2.768 (14)	144 (3)

Symmetry codes: (i) $-x, y, 1-z$; (ii) $x, 1+y, z$; (iii) $-x, y, -z$; (iv) $-x, 1+y, 1-z$.

The displacement parameters of the terminal methyl group on the disordered methionine side chain were constrained using the ISOR instruction (isotropic approximation) in *SHELXTL* (Sheldrick, 1997b). Difference peaks for the H atoms on two of the water molecules were located. The positions of these H atoms were refined with the O—H and H \cdots H distances restrained to reasonable values.

Data collection: *SMART* (Bruker, 1999); cell refinement: *SMART*; data reduction: *SAINTE* (Bruker, 2000); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1990); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997a); molecular graphics: *SHELXTL* (Sheldrick, 1997b); software used to prepare material for publication: *SHELXL97*.

This research was supported in part by the National Institute for Drug Abuse (NIDA) and the Office of Naval Research (ONR). Portions of this study were conducted at the Cornell High Energy Synchrotron Source (CHESS), which is supported by the National Science Foundation under award DMR 97-3424. Use of the Macromolecular Diffraction facility (MacCHESS), is supported by award RR-01646 from the National Institutes of Health, through its National Center for Research Resources.

References

- Bruker (1999). *SMART*. Version 5.059. Bruker AXS Inc., Madison, Wisconsin, USA.
- Bruker (2000). *SADABS* (Version 2.01) and *SAINTE* (Version 6.02A). Bruker AXS Inc., Madison, Wisconsin, USA.
- Deschamps, J. R., C. George, C. & Flippen-Anderson, J. L. (1996). *Peptide Science (Biopolymers)*, **40**, 121–139.
- Doi, M., Ishida, T., Inoue, M., Fujiwara, T., Tomita, K., Kimura, T. & Sakakibara, S. (1984). *FEBS Lett.* **170**, 229–231.
- Doi, M., Tanaka, M., Ishida, T., Inoue, M., Fujiwara, T., Tomita, K., Kimura, T., Sakakibara, S. & Sheldrick, G. M. (1987). *J. Biochem.* **101**, 485–490.
- Flack, H. D. (1983). *Acta Cryst.* **A39**, 876–881.
- Frederickson, R. C. A., Smithwick, E. L., Shuman, R. & Bemis, K. G. (1981). *Science*, **211**, 603–605.
- Griffin, J. F., Langs, D. A., Smith, G. D., Blundell, T. L., Tickle, I. J. & Bedarkar, S. (1986). *Proc. Natl Acad. Sci. USA*, **83**, 3272–3276.
- Karle, I. L., Karle J., Mastropalo D., Camerman, A. & Camerman, N. (1983). *Acta Cryst.* **B39**, 625–637.
- Sheldrick, G. M. (1990). *Acta Cryst.* **A46**, 467–473.
- Sheldrick, G. M. (1997a). *SHELXL97*. University of Göttingen, Germany.
- Sheldrick, G. M. (1997b). *SHELXTL*. Version 5.10. Bruker AXS Inc., Madison, Wisconsin, USA.
- Spek, A. L. (2001). *PLATON*. Utrecht University, The Netherlands.